# The influence of air pollution on moss - dwelling animals: 1. Methodology and composition of flora and fauna

## Werner A. STEINER

Swiss Federal Research Station for Arboriculture, Viticulture and Horticulture, Department of Entomology and Nematology, CH-8820 Wädenswil.

The influence of air pollution on moss-dwelling animals: 1. Methodology and composition of flora and fauna. - This paper is part of a study investigating the suitability of the moss-dwelling fauna to act as a monitoring system for air pollution effects. The strategy developed to study the influence of air pollution on the fauna is introduced and ecological aspects of moss-invertebrate associations are presented. An attempt is made to characterise the flora and fauna as encountered in mosses from tiles and walls. Finally, the overall floristric and faunistic composition are given (including a complete list of species).

**Key-words:** Moss-dwelling animals - Air pollution - Moss species - Moss-invertebrate associations.

## INTRODUCTION

The survey of pollution levels and of their effect on the biosphere is necessary to prevent injuries both to ecosystems and to human health. The physical and chemical monitoring of a few important pollutants is already well established in environmental surveys. Single species' tests are the major tools for estimating damage from environmental stress (CAIRNS 1983). However, the degree of reliability to predict response at the community level is relatively small (OKKERMAN *et al.* 1991). To allow accurate predictions, the study of ecosystem disturbances should be based on a multispecies approach (including bioindicators, sensu ARNDT *et al.* 1987) and on the lowest possible taxonomic level.

Ecological monitoring has been poorly developed, although it is successfully used to indicate water quality ("Saprobiensystem" described by KOLKWITZ 1959) and specific air pollutants (MUKAMMAL 1976; ARNDT *et al.* 1987). Lichens and mosses, for example, are often used in regional surveys to indicate SO<sub>2</sub> pollution (e.g. LE BLANC & DE SLOOVER 1970; HECK & BRANDT 1977). No equivalent monitoring system exists

Manuscript accepted 14.01.1994.

for soil ecosystems. Knowledge about terrestrial invertebrates as indices of environmental quality is alarmingly poor (SCHUBERT 1985; ARNDT *et al.* 1987). As the soil and its associated fauna are very complex, the use of simply structured, but similarly composed communities may provide an alternative for studying pollution effects on soil ecosystems.

Animal communities of moss cushions could be used for this purpose. The moss-dwelling fauna has a close similarity with the fauna of soils (NICHOLAS 1975). Epilithic (living on or among stone) moss cushions are abundant in urban as well as in rural environments (GILBERT 1968), thus allowing for comparative studies. An advantage of the moss-dwelling fauna is its availability for experimental purposes. Tegulous (living on tiles; from Latin: tegula = tile) moss cushions, for example, can easily be manipulated in the lab (e.g. fumigation with gaseous air pollutants) or transplanted to sites with different environmental characteristics.

The suitability of the moss-dwelling fauna to act as a monitoring system for air pollution effects was studied near Zürich from 1982 to 1989. The study involved three main approaches: 1) survey of natural communities in epilithic moss cushions, 2) fumigation (with  $SO_2$ ) and 3) exposure of tegulous moss-invertebrate associations to different levels of air pollution in the field. The present paper introduces biological aspects of moss-invertebrate associations. It then describes the strategy developed to study effects of air pollution on the moss-dwelling fauna. Finally, the flora and fauna of both epilithic and tegulous moss cushions is characterised, and a list of species is presented.

## **MOSS-INVERTEBRATE ASSOCIATIONS**

### 1. THE MOSSES AND THEIR ASSOCIATED FLORA

Mosses can be classified into three groups according to their moisture demands (RAMAZZOTTI 1972). In man-made environmental regimes, mosses of RAMAZZOTTI's groups "intermediate" and "dry" find a large variety of artificial substrata to colonise (SEAWARD 1979). Thus, only "intermediate" and "dry" mosses are considered in the present study, while the third group ("moist" mosses) is disregarded. The "moss cushion microcosm" comprises an autotrophic (i.e. algae, lichens and liverworts) and a heterotrophic microflora (i.e. bacteria, yeasts, and filamentous fungi). The latter is responsible for the primary decomposition of organic matter in analogy to decomposition in the soil ecosystem (DAVIS 1981). Thus, it is possible that both the moss and its associated microflora influence the moss-dwelling fauna.

### 2. The moss-dwelling fauna

Invertebrates associate with mosses wherever these grow (e.g. STAFANSKI 1923; TRAVÉ 1963; NIELSEN 1967; MCINNES 1991). According to GERSON (1982), the moss-invertebrate community includes the following taxa: Protozoans, rotifers, nematodes, tardigrades, and arthropods (especially mites and insects). TRAVÉ (1963)

pointed out that there are two more or less mutually exclusive faunas on mosses: Protozoans, rotifers, nematodes and tardigrades belong to the aquatic, arthropods to the terrestrial fauna.

The fauna of moss cushions is affected by a large number of interrelated abiotic and biotic factors. The moisture content of the microhabitat, probably along with the amount of insolation, seems to be the most important factor influencing the moss-dwelling fauna (TRAVÉ 1963; RAMAZZOTTI 1972). General aspects of the surroundings (open or covered areas), topographical location (height above ground, inclination), type of wall (sunken, freestanding or building), solar radiation and temperature are thus important as they regulate the water supply and/or evaporation (NIELSEN 1967; BERTRAND 1975). The moss-dwelling fauna does not appear to be specialised on any particular moss species (RAMAZZOTTI 1972; PÉREZ-IÑIGO 1975; GERSON 1982; KATHMAN & CROSS 1991). This may be partly understood by considering the moss cushion as a suitable microenvironment for the microflora and microfauna rather than as a main food source. Human activities may affect the bryofauna directly (e.g. release of animal toxic compounds) as well as indirectly (e.g. by releasing phytotoxic compounds, or by offering artificial substrata such as walls and roofs).

## MATERIAL AND METHODS

### **1. GENERAL STRATEGY**

In accordance with DIAMOND (1986), the present study included the following approaches: a) a survey of natural communities (faunistic survey), b) exposure experiments (field experiments), and c) fumigation experiments with SO<sub>2</sub> (laboratory experiments). The faunistic survey carried out simultaneously at all sites (main faunistic survey) allowed the comparison of natural moss-dwelling animal communities subjected to different air pollution levels (STEINER 1994, 1995*a*). The reliability of community analysis (terrestrial fauna) based on one single sampling was assessed by a second faunistic survey, carried out partly in the same locations (STEINER 1995*a*). The reliability of results on the aquatic fauna was estimated by investigating long-term dynamics (1984-1989) as well as annual fluctuation (1983-1985) of nematode and tardigrade populations (STEINER 1995*b*). In the exposure and fumigation experiments invertebrate communities of tegulous mosses were exposed to different levels of air pollution in the field (tab. 1, STEINER 1995*c*), and to different SO<sub>2</sub> levels in fumigation chambers, respectively (STEINER 1995*c*).

The following taxa were considered: Rotifers, nematodes and tardigrades as representatives of the aquatic fauna, mites (with emphasis on oribatid mites), insects (with emphasis on springtails) and other arthropods as representatives of the trerrestrial fauna. Identification to species level was carried out for nematodes, tardigrades, oribatid mites and springtails. Rotifers were analysed as a group because their determination is difficult (J. Donner, pers. comm.). Moss species were partially identified by E. Urmi and collaborators (Institut für Systematische Botanik).

#### WERNER A. STEINER

#### 2. STUDY SITE DESCRIPTION

The study was conducted at urban sites in the city of Zürich and at rural sites in the canton of Zürich (tab. 1). "Study site" refers to an ancient or actual air pollution recording site, as well as to its direct surroundings, expected to have about the same pollution levels as the recording site itself. All study sites are situated at altitudes between 400 and 550 m above sea level, except site "A", which lies at an elevation of 800 m.

#### Tab. 1

Annual mean values of gaseous air pollutants at the study sites (SO<sub>2</sub>, NO and NO<sub>2</sub> in  $\mu$ g/m<sup>3</sup>, CO in mg/m<sup>3</sup>; Abbr. = abbreviations for study sites, as used in text and figures; Year = period of air sampling; Type of study: fs = faunistic survey of October 1984, s = supplementary faunistic survey of June 1987, e = exposure experiments, dyn = study of seasonal and long-term population fluctuations, fum = fumigation experiments; urb. cat. = urbanisation categories: uht = urban high traffic, ult = urban low traffic, rlt = rural low traffic; - : not measured).

Study sites			Туре					urb.
	Abbr.	Year	of study	SO <sub>2</sub>	CO	NO	NO <sub>2</sub>	cat.**
Rosengartenstrasse	R	82/83	fs, s, e	47	4.4	357	87	uht
Weststrasse	W	82/83	fs, s	50	5.6	236	79	uht
Stampfenbachstrasse	Т	82/83	fs, s, e	44	2.5*	157	54	uht
Überlandstrasse	U	82/83	fs, s	28	2.4	166	66	uht
Büchnerstrasse	В	82/83	fs, s	43	1.6	54	56	ult
Eglistrasse	E	82/83	fs, s	29	1.4	58	53	ult
Kaserne	K	82/83	fs, s, e	45	1.4	45	56	ult
Seefeldstrasse	S	82/83	fs, s	31	1.2	51	50	ult
Bachtel	А	84/86	fs	12	0.3	6	15	rlt
Höri Berg	H	84/86	fs	7	0.4	22	28	rlt
Männedorf	Μ	84/86	fs	15	0.4	16	30	rlt
Oberstammheim	0	84/86	fs	12	0.3	9	28	rlt
Rämistrasse	Rä	82/83	e	35	6.1	217	64	uht
Forchstrasse*	F	82/83	e	31	1.9	93	62	uht
Clausiusstrasse	С	84/85	e	43	1.9	44	61	ult
Mythenquai	Y	82/83	e	28	1.2	66	48	ult
Unterer Letten*	L	82/83	e	26	1.1	54	48	ult
Letzibad*	Ζ	82/83	е	27	1.2	50	50	ult
Karl Staufferstrasse*	KS	82/83	e	26	1.0	36	43	ult
Dübendorf	D	82/83	е	21	1.3	48	55	rlt
Tänikon	Tä	85	e	12	-		21	rlt
Hohenbühlstrasse*	Dyn	82/83	dyn	36	3.4	178	66	uht
Birmensdorf	Bi	84/85	fum	12	-	17	26	rlt

Levels of gaseous pollutants at the sites "D", "Tä" and "K" were continually monitored by the Swiss Federal Laboratory for Material Testing and Research (in Dübendorf), and at site "Bi" by BLEULER & BUCHER (1984). Mean values for the sites A", "H", "M", and "O" were determined by a commercial institute of applied ecology (Oekoscience, Zürich). Pollution values for site "C" were given by HUTER (1986) and for sites other than those mentioned above by the Air Pollution Control Agency of Zürich (BACHMANN-STEINER *et al.* 1983).

\*Annual mean values are estimated based on measurements and guidelines given by the Air Pollution Control Agency of Zürich (BACHMANN-STEINER *et al.* 1983).

\*\*Urbanisation categories are defined according to traffic volume (BACHMANN-STEINER *et al.* 1983) and distance from the center of the city.

#### 3. SAMPLING AND SAMPLE PROCESSING

Sampling for the main faunistic survey was carried out in October 1984, for the second (supplementary) survey in June 1987, and for the study of population fluctuations between 1983 and 1989. Fumigation and exposure experiments were performed between 1982 and 1985.

The sampling unit size used for the analysis of the aquatic fauna was fixed empirically. As a general rule, a sampling unit should measure at least 20 times the length of the animals to be collected (COCHRAN 1963). A sampling unit with a diameter of 11 mm (cores of ca. one cm<sup>2</sup>) for the aquatic fauna, and 25 mm (cores of five cm<sup>2</sup>) for the terrestrial fauna was considered to satisfy this criterion and was used throughout the study.

The influence of moss cushion characteristics (e.g. size, thickness, orientation) and of the season on the aquatic fauna had been analysed in preliminary studies (STEINER 1990). In addition, the distribution of nematode and tardigrade species was investigated at different strata of the habitat (i.e. within moss cushions, within walls, and within sites). Sampling plans were then defined according to the aims of the different approaches (STEINER 1994, 1995*a*, *b*, *c*).

Samples for both the aquatic and the terrestrial fauna were taken from the same moss with cylindrical steel corers and passed immediately into glass vials for transport to the laboratory. The following sample sizes were used: 1) main and supplementary faunistic survey: four cm<sup>2</sup> for the aquatic fauna, ten cm<sup>2</sup> for the terrestrial fauna; 2) dynamics of the aquatic fauna: five cm<sup>2</sup>; 3) fumigation and exposure experiments: one cm<sup>2</sup>.

Extraction of the fauna started generally one day after sampling. The aquatic fauna was extracted using a modified version of OOSTENBRINK's funnel-spray method (STEINER 1990). Moss cores were moistened prior to extraction and were then mechanically macerated. After an extraction time of three days, the moss samples were analysed under a dissection microscope for remaining organisms. The extracted animals were killed in hot water ( $60^{\circ}$ C for two min). After a minimum preservation time of one week in TAF (i.e. 14 parts formol 38%, four parts triethanolamine; 82 parts *Aq. dest.*), the total number of extracted animals was determined under a dissection microscope (magnification 40 x). Nematodes and tardigrades were dehydrated using the rapid method of SEINHORST (1959). A maximum of 300 randomly selected individuals per sample was permanently mounted in glycerine. Unidentified individuals were proportionally distributed to the taxa found in that sample.

Arthropods were extracted in a modified MACFADYEN extractor (BIERI *et al.* 1978), gradually raising the temperature from 22 to  $32^{\circ}$ C during the extraction time of six days. Samples were moistened by spraying water every 12 hours. For the last four hours the temperature was increased to  $50^{\circ}$ C. The extracted arthropods were collected in isopropanol and counted under a dissecting microscope (magnification 40 x). Macroarthropods could be identified directly, whereas microarthropods had to be pre-treated either with lactic acid or KOH. For identification, mites were temporarily

mounted in cavity slides. The immature stages of the Oribatulidae (Oribatida) were processed separately (STEINER 1989). Springtails were either mounted permanently in Hoyer (Berlese) medium, or for quick observation simply placed in cavity slides.

## 4. DATA PROCESSING

Whenever possible, unidentified individuals (mainly preimaginal forms) in a sample were assigned to related single taxa (STEINER 1989) or were proportionally distributed on groups of closely related taxa occurring in the same sample. Species with uncertain taxonomic status were treated as follows: the nematodes *Plectus acuminatus* Bastian and *P. cirratus* Bastian as *P. acuminatus s.l.*; *Plectus parietinus* Bastian and *P. silvaticus* Andrássy as *P. cf. parietinus*; juveniles of the closely related species of the genus *Plectus*, as well as first larval instars of *Chiloplectus cf. andrassyi* (Timm) as "*Plectus* (larvae; undetermined)"; the Neotylenchidae as different taxa, although the *Neotylenchidae sp. 1* and *Neotylenchidae sp. 2* consisted only of larvae, the remaining taxa of adults; the tardigrades *Hypsibius convergens* (Urbanowicz) and *H. dujardini* (Doyère) as *H. convergens s.l.*; the oribatid mites of the genus *Scutovertex* as *S. cf. sculptus* Michaël, a species typical of the habitat considered in this study (S. Woas, pers. comm.).

The sign test was applied to compare the ratio of higher taxonomic categories (main taxa) in epilithic and tegulous moss cushions.

## FLORA

Species composition of the samples (epilithic mosses) taken in the main faunistic survey of 1984 is presented in tab. 2. More than 90% of the moss species belong to the Dicrananae, a superorder comprising mosses of acrocarpous growth form. *Bryum argenteum* Hedw. and *Tortula muralis* Hedw., are the most frequent species, accounting for 44% of the total number of samples. According to GILBERT (1971), the dominance of *B. argenteum* indicates both polluted conditions and a high nitrogen supply. Many of the species listed in tab. 2 seem to be typical of urban stonework, and are ubiquitous species (GILBERT 1971). The study of the fluctuations of the aquatic fauna (STEINER 1994) was confined to a large moss cushion of the species *Homalothecium cf. sericeum* (Hedw.).

Tegulous moss species sampled during fumigation and exposure experiments (STEINER 1995c) were dominated by the cosmopolitan species *Hypnum cupressiforme* Hedw. *s.l.*, *Tortula ruralis* Gärtn. Meyer & Scherb., *Bryum capillare* Hedw. *s.l.* and *Caratodon purpureus* (Hedw.). A very similar flora is found in roof habitats in England (GILBERT 1968) and Germany (VON DER DUNK 1988).

## FAUNA

## 1. GENERAL ASPECTS

The present survey is the first comprehensive study of the fauna of epilithic mosses in Switzerland. In the past, surveys were either restricted to one or two main

#### Tab. 2

Overall species composition of moss samples taken from tiles during fumigation and exposure experiments (102 and 393 samples, respectively), and during the survey of natural communities carried out in October 1984 (216 samples; epilithic mosses). Systematic categories are according to FRAHM & FREY (1983).

Moss species	Relative ab	oundance (%)
-	Tegulous	Epilithic
SUPERORDER	mosses	mosses
Ceratodon purpureus (Hedw.)	16.4	6.5
Encalypta streptocarpa Hedw.		0.5
Tortula intermedia (Brid.)		0.9
Tortula muralis Hedw.		19.4
Tortula ruralis Gärtn. Meyer & Scherb.	16.6	
Barbula fallax Hedw.		0.9
Barbula hornschuchiana Schultz		0.5
Barbula rigidula (Hedw.)		12.0
Barbula sp.		0.5
Schistidium apocarpum (Hedw.)	8.4	11.6
Bryum argenteum Hedw.		25.0
Bryum capillare Hedw. s.l.	16.4	6.9
Bryum sp.		2.3
Orthotrichum anomalum Hedw.		1.4
Orthotrichum diaphanum Brid.		2.8
DICRANANAE $\sum$ acrocarpous mosses	57.4	91.2
Pseudoleskeella catenulata (Schrad.)		0.5
Amblystegium serpens (Hedw.)		6.5
Homalothecium sp.		0.5
Brachythecium sp.	14.1	
Rhynchostegium sp.		0.5
Hypnum cupressiforme Hedw. s.l.	28.5	0.5
Pylasia polyantha (Hedw.)		0.5
<b>HYPNANAE</b> $\sum$ pleurocarpous mosses	42.6	8.8

taxa, and/or the epilithic habitat was sampled only casually (AMMANN 1908; HEINIS 1908, 1910; GISIN 1943; SCHWEIZER 1949, 1951, 1956; BARTOS 1950; ALTHERR 1952; PSCHORN-WALCHER & GUNHOLD 1957).

A total of more than 230 taxa and close to 240'000 individuals was recorded (appendix k). Nematodes and tardigrades accounted for 75% of the total number of individuals. Diversity was highest within nematodes (56 taxa), followed by mites (47; ticks included, oribatid mites excluded), springtails and oribatid mites (39 each), arthropods (37; previous taxa excluded) and tardigrades (16). As low density species were by far more numerous than high density species, additional sampling would certainly reveal other species. Significant differences exist between the faunal composition of mosses from tiles and from walls. Among the aquatic fauna, nematodes dominate in epilithic mosses (sign test, n=13, P < 0.05), whereas tardigrades prevail in tegulous mosses (sign test, n=16, P < 0.01). Among the arthropods, mites (oribatid mites included) dominate quantitatively in mosses from walls (sign test, n=13, P < 0.01), whereas there was no significant trend for the ratio mites/springtails in mosses from tiles.

#### MAXIMUM DENSITY OF HIGHER TAXONOMIC CATEGORIES

#### Tab. 3

Maximum density values (number of individuals/cm<sup>2</sup>) of higher taxonomic categories in tegulous and epilithic moss samples (Ori: oribatid mites).

Taxon	Tegulous mosses	Epilithic mosses	
Rotifers	192	97	
Nematodes	354	879	
Tardigrades	927	227	
Springtails	53	388	
Oribatid mites	15	89	
Mites (Ori excluded)	2	30	
Arthropods*	8	3	

\* Taxa above excluded.

Tab. 3 shows the maximum density of main taxa for tegulous and epilithic moss cushions. Maximum densities of moss-dwelling rotifers (NIELSEN 1967; WRIGHT 1991), nematodes (NIELSEN 1967) and tardigrades (RAMAZZOTTI 1972; MORGAN 1977) approximate those recorded in the present study. The density of moss-dwelling springtails was several times higher in the present study than is reported in the literature (BENGTSON *et al.*, 1974, as cited by GERSON, 1982).

Comparison of the moss-dwelling fauna (tab. 3) with the fauna of soil (PETERSEN & LUXTON 1982) shows similar density values for rotifers and mites. Values reported by PETERSEN & LUXTON (1982) for tardigrades (10 individuals/cm<sup>2</sup>) and springtails (70 individuals/cm<sup>2</sup>) are smaller than corresponding values in tab. 3, whereas the density of soil-dwelling nematodes (3'000 individuals/cm<sup>2</sup>) is about three times larger. Taking into account that soil samples are usually thicker than the moss samples (e.g. 1 - 2 cm in the present study), moss-dwelling invertebrates occur at higher densities (at least temporally) than soil-dwelling animals.

### 2. Species composition of the aquatic fauna

#### Rotifers

Rotifers were recorded in all epilithic moss cushions, as well as in all moss cushions on tiles, before the fumigation and exposure of the tiles started. The rotifers found in the present study belong mainly to the Bdelloidea. This group is especially abundant in bryophytes of the Temperate Zone (J. Donner, pers. comm.), and many bdelloid species have a cosmopolitan distribution (HYMAN 1951; DONNER 1965).

### Nematodes

In tegulous mosses, bacterial feeders (i.e. Monhysterida, Araeolaimida, Rhabditida) were more abundant than predators such as *Paratripyla intermedia* (Bütschli) and *Prionchulus muscorum* (Dujardin), fungivorous (e.g. *Aphelenchoides*, Tylenchidae, and Neotylenchidae) and omnivorous nematodes (Dorylaimidae).

Chiloplectus cf. andrassyi was the most abundant species, followed by *P. intermedia*, other species of the family Plectidae (Araeolaimida), *Tylocephalus auriculatus* (Bütschli) and *P. muscorum* (appendix b). In epilithic mosses, fungivorous nematodes were predominant with Aphelenchoides sp. 5 being the most abundant species, except at site "Dyn". The high number of bacterial feeders in tegulous mosses and of fungivorous nematodes in epilithic mosses indicates that the microflora of the two habitats differs substantially.

Many of the numerically dominant species were also characterised by high frequencies in tegulous and/or in epilithic moss cushions (tab. 4). *Plectus acuminatus s.l.*, *P. parietinus*, *P. parvus* Bastian, *P. intermedia*, and *P. muscorum* seem to be ubiquitous (STEINER 1990). For *C. cf. andrassyi*, *Aphelenchoides sp. 4*, and *Aphelenchoides sp. 5* the actual distribution is unknown.

Nematodes are amongst the most numerous animals feeding on primary decomposers. By grazing on the microflora, nematodes can have considerable impact on the distribution and activities of bacteria and fungi (SCHIEMER 1982; ANDERSON 1988) and participate thus indirectly in the decomposition of dead plant material in the moss cushions.

Moss-dwelling taxa frequently encountered (rf > 20%) either in tegulous (first sampling; 74 samples) or epilithic moss cushions (survey of natural communities; Ne and Ta: 120 samples, Ar: 96). (rf: relative frequency; Ne, Ta, Ar: nematodes, tardigrades, arthropods; - : not present).

		Tegulous	Epilithic
		mosses	mosses
Taxon		rf (%)	rf (%)
Ne	Plectus acuminatus Bastian s.l.	43	12
	Plectus cf. parietinus Bastian	16	38
	Plectus parvus Bastian	26	34
	Chiloplectus cf. andrassyi (Timm)	74	22
	Aphelenchoides sp. 4	3	32
	Aphelenchoides sp. 5	1	46
	Paratripyla intermedia (Bütschli)	58	18
	Prionchulus muscorum (Dujardin)	32	26
Га	Macrobiotus areolatus Murray	69	. 4
	Macrobiotus cf. artipharyngis Iharos	50	-
	Macrobiotus hufelandi Schultze	78	44
	Macrobiotus persimilis Binda & Pilato	1	39
	Hypsibius convergens Urbanowicz s.l.	28	30
	Hypsibius oberhäuseri (Doyère)	27	30
	Milnesium tardigradum Doyère	3	36
	Echiniscus blumi Richters	68	2
Ar	Camisia segnis (Hermann)	1	34
	Tectocepheus sarekensis Trägårdh	19	81
	Scutovertex cf. sculptus Michaël	24	8
	Zygoribatula exilis (Nicolet)	-	39
	Trichoribates cf. trimaculatus (Koch)	-	52
	Eupodes sp.	-	62
	Tydeus sp.	-	35
	Bdella longicornis (L.)	-	23
	Polyxenus lagurus L.	-	31
	Xenylla brevisimilis Stach	31	9
	Chironomidae sp.	-	27
	Psychodidae sp.	-	27

Tab. 4

## Tardigrades

The Eutardigrada represented the major class of the phylum Tardigrada (appendix c). This is mainly due to the sampling plan having been restricted to northerly exposed habitats. Heterotardigrada occur preferentially in moss cushions exposed to direct solar radiation (RAMAZZOTTI 1972), which agrees with the high frequency of *Echiniscus blumi* Richters in tegulous moss cushions (tab. 4). *Macrobiotus hufelandi* Schultze was the most common tardigrade found in the present study (tab. 4, appendix c).

Most tardigrades listed in tab. 4 are cosmopolitan species (MAUCCI 1986). *Milnesium tardigradum* Doyère and certain *Macrobiotus* species found in this study (i.e. *M. areolatus* Murray, *M. hufelandi*, *M. persimilis* Binda & Pilato, and *M. richtersi* Murray) are predators of the aquatic fauna. It is believed, however, that some *Macrobiotus* species are actually non specialised feeders. For instance, individuals of *M. hufelandi* were observed to ingest filamentous algae. *Macrobiotus cf. artipharyngis* Iharos, as well as *Hypsibius* spp. probably ingest bacteria and/or cell contents of algae, fungi or moss leaves.

#### 3. SPECIES COMPOSITION OF THE TERRESTRIAL FAUNA

Arthropods were the most diverse phylum of the terrestrial fauna, but poorer in numbers than representatives of the aquatic fauna (appendix k). Four classes of arthropods (i.e. arachnids, crustaceans, myriapods, and insects) were encountered (appendix d to i). Annelids and molluscs were extremely rare in the present study and are disregarded.

## Arachnids

Araneae, Pseudoscorpiones, Opiliones and Acari are the four orders of the class Arachnida recorded in the present study (appendix d to g). The order Acari is the largest group represented by the suborders Gamasida, Ixodida, Actinedida, and Oribatida. While the presence of Ixodida in the moss habitat was unexpected, representatives of the other suborders are known to dwell in mosses (GERSON 1969, 1982; GERSON & SEAWARD 1977).

Oribatid mites reached the highest numbers of all arthropod taxa (appendix e). Most of the recorded oribatid species are microphytophages or non-specialised feeders with holarctic to world-wide distribution (WEIGMANN & KRATZ 1981). Oribatulidae (nine species) and Tectocepheidae (one species) were the numerically dominant families, while representatives of primitive oribatid families (supercohort Macropylides) were rare. Within the oribatid mites, five species are typical dwellers of epilithic or tegulous moss cushions (tab. 4). The most frequent and numerically dominant species in epilithic mosses (main survey of 1984) was *Tectocepheus sarekensis* Trägårdh (tab. 4, appendix e), closely related to *T. velatus* Michaël (KNÜLLE 1954). The latter species is one of the most common mite species occurring

in the soil, and is believed to reach greatest numbers in dry habitats (PSCHORN-WALCHER & GUNHOLD 1957; MURPHY & JALIL 1964). The exclusive occurrence and high frequency of T. sarekensis in mosses from tiles and walls (appendix e, tab. 4) can be explained by its higher resistance to desiccation as compared with T. velatus (JALIL 1972). Along with T. sarekensis, Zygoribatula exilis (Nicolet) was the most abundant mite species in epilithic moss cushions (appendix e). This agrees with the composition of epilithic and corticolous (i.e. living on bark) mosses in the region of Zürich (PSCHORN-WALCHER & GUNHOLD 1957) and in several European countries (STRENZKE 1952; PSCHORN-WALCHER & GUNOLD 1957; TRAVÉ 1963). The relative abundance of Trichoribates cf. trimaculatus (Koch) and Camisia segnis Hermann was low (appendix e). Nevertheless these species were frequently encountered in epilithic moss cushions (tab. 4). While T. cf. trimaculatus is known to dwell in this habitat (STRENZKE 1952; PSCHORN-WALCHER & GUNHOLD 1957), C. segnis is usually found in corticolous lichens (LEBRUN 1976; ANDRÉ et al. 1984). Scutovertex cf. sculptus was the only oribatid species in tegulous mosses with a relative frequency > 20% (tab. 4). Its related species S. minutus (Koch) seems to be typical of roof habitats (STRENZKE 1952; SMRZ 1992). According to S. Woas (pers. comm.), the two species are likely to be mixed up, and notes on their ecology are probably not very specific.

Gamasid mites (nine families) only occurred at low population densities (appendix f), with Asca aphioides (L.) the predominant species. Several of the recorded gamasid species prey upon diverse bryophagous taxa (WALLWORK 1967; GERSON 1972), including members of the aquatic fauna, enchytraeids, mites, as well as on eggs and larvae of higher insects.

Actinedid mites are usually represented in moss cushions by numerous species (SCHWEIZER & BADER 1963), however, their ecology is almost unknown (ANDRÉ 1986). In the present study, actinedid mite numbers (appendix g) were lower than those of oribatid mites (appendix e), but higher than those of gamasid mites (appendix f), complying with findings by ANDRÉ (1975) for the fauna of corticolous lichens. Among actinedid mites, three families (Eupodidae, Tydeidae, and Bdellidae) seem to be typical of epilithic moss cushions (tab. 4). Eupodid mites were the second most frequent arthropods in this habitat. They probably prey on other mites as indicated by KRANTZ (1978) for an eupodid species dwelling in mushroom beds. According to H. André (pers. comm.), at least two different *Eupodes sp.* were sampled in the present study. The Tydeidae were the most diverse actinedid family (appendix g). This agrees again with findings forthecorticolous fauna (ANDRÉ 1986). Tydeid mites probably prey on nematodes (SANTOS et al. 1981), which were very abundant in epilithic moss cushions of the present study (appendix b). The Bdellidae were dominated by the species Bdella longicornis (L.). This predatory species was common in epilithic mosses (tab. 4, appendix g) and was also found in terrestrial moss cushions (SCHWEIZER 1951). Predators like Bdellidae and Rhagidiidae (appendix g) probably form an important link between soft bodied oribatid mites on which they feed, and the larger carnivorous arthropods (WALLWORK 1967). As the latter group was scarce in epilithic and tegulous mosses (appendix d), the importance of predatory actinedid mites increases in these habitats.

### Crustaceans

Only two copepods were found in epilithic mosses taken for preliminary studies. Malacostracans (i.e. isopods) were more abundant (appendix d). They were usually found under the moss cushions, using the moss as a refuge. According to GERSON (1982), copepods and malacostracans usually associate with aquatic mosses.

### **Myriapods**

Chilopods and diplopods were collected in epilithic (appendix d) as well as in corticolous mosses (PSCHORN-WALCHER & GUNHOLD 1957). Predatory chilopods (i.e. Lithobiidae) and diplopods (other than *Polyxenus lagurus* L.) are probably occasional guests, while *P. lagurus* reached a relatively high frequency (tab. 4). According to DUNGER (1983), the latter species is typical of the corticolous habitat.

## Insects

The Apterygota are usually more numerous in bryophytes than the Pterygota, with springtails representing the dominant group (GERSON 1982). In the present study, a total of 36 collembolan species of the suborder Arthropleona and three species of the suborder Symphypleona were recorded, most of them in epilithic moss cushions (appendix h). A similar species composition was found by GISIN (1943) on bark. Xenylla brevisimilis Stach was along with S. cf. sculptus the only arthropod species typical of tegulous mosses (tab. 4). According to ANDRÉ (1983), X. brevisimilis is frequently encountered in foliose lichens on bark. The high abundance of Schoettella ununguiculata (Tullberg) in the faunistic survey of 1984 is based on a single sample with an exceedingly high density (388 individuals/cm<sup>2</sup>), but frequency of this species was low (8%). Many species of springtails are non-specific feeders (HALE 1967; VISSER 1985), ingesting dead plant matter, bacteria and fungi (PETERSEN & LUXTON 1983). Since enchytraeids, earthworms and large diplopods are usually absent from moss cushions, the contribution of springtails in particular and arthropods in general to the decomposition of organic material is far greater in mosses than in soil. By their high motility, springtails also play an important role in the dispersal of the microflora (CHRISTEN 1975; PETERSEN & LUXTON 1982).

The Pterygota were represented by ten insect orders (appendix i). Dipterans were the most conspicuous holometabolous insects in epilithic mosses (tab. 4, appendix i). They represent the insect order most intimately associated with bryophytes, though some taxa of springtails show a similar degree of association (GERSON 1982). Larvae of many dipteran families, including Chironomidae, Psychodidae and Cecidomyiidae, are known to dwell in bryophytes (BRAUNS 1976; GERSON 1982).

## CONCLUDING REMARKS

Although epilithic and tegulous mosses are climatically extreme habitats (TRAVÉ 1963; RAMAZZOTTI 1972), the inhabiting fauna was surprisingly rich in taxa.

Based on the relationship between sample size and species number (the probability of a species' presence increases with sample size), and taking into account that several taxa were ignored in the present study, about 350 to 600 species can be expected to be found in dry mosses of central Europe (STEINER 1990). The high diversity (appendix a-i) as well as the high densities of moss-dwelling populations (tab. 3) indicate that mosses provide – at least temporally – favourable conditions for the associated fauna.

The wide ranging distribution of several taxa is probably a result of their mechanism of dissemination. Moss-dwelling animals evolved different adaptive life forms (eggs, cysts, tuns) to resist desiccation. These may be blown away by wind and can potentially reach any suitable habitat. Therefore, differences between mosses from roofs and walls (tabs. 3 and 4, appendix), or between study sites of a regional survey (STEINER 1994, 1995*a*) are unlikely to originate from limits to dispersal. The influence of air pollution, season and/or other environmental factors on moss-dwelling invertebrate communities is investigated by STEINER (1994, 1995*a*, *b*, *c*).

#### ACKNOWLEDGEMENT

The present paper was part of a doctoral thesis carried out under Prof. V. Delucchi. The project was financially supported by the Swiss National Science Foundation (grant no. 5.521.330.817/3). Realisation of this work was only possible thanks to the support provided by Prof. V. Delucchi and the following taxonomists: Dr. E. Urmi (moss species), Prof. A. Zullini and Dr. J. Klingler (Nematoda), Dr. J. Travé (Oribatida), Dr. H. André (Actinedida), Mme I. Bals (Gamasida), Prof. R. Jordana (Collembola), Dr. J. Walter (Aranea), and Prof. W. Sauter (Diptera). The author is grateful to the Air Pollution Control Agency of Zurich and to the Swiss Federal Laboratory for Material Testing and Research (Dübendorf) for allowing the exposure of moss covered tiles in proximity to their pollution gauges and for making pollution ratings available. A special thank is extended to Dr. K. Tschudi-Rein for her corrections on the manuscript.

#### REFERENCES

- ALTHERR, E., 1952. Les nématodes du Parc national suisse, 2'. Ergebnisse der wissenschaftlichen Untersuchungen im schweizerischen Nationalpark 26, 315-356.
- AMMANN, J., 1908. Beitrag zur Kenntnis der schweizerischen Tardigraden. Diss. Univ. Bern.
- ANDERSON, J. M. 1988. Spatiotemporal effects of invertebrates on soil processes. *Biol. Fertil.* Soils 6, 216-227.
- ANDRÉ, H. M., 1975. Observations sur les acariens corticoles de Belgique. Fondation Universitaire Luxembourgeoise, Sér. Notes de Recherche 4, 5-30.
- ANDRÉ, H. M., 1983. Notes on the ecology of corticolous epiphyte dwellers. 2. Collembola. *Pedobiologia* 25, 271-278.
- ANDRÉ, H. M., 1986. Notes on the ecology aof corticolous epiphyte dwellers. 4. Actinedida (especially Tydeidae) and Gamasida (especially Phytoseiidae). Acarologia 27(2), 107-115.
- ANDRÉ, H. M., PH. LEBRUN & S. LEROY, 1984. The systematic status and geographical distribution of *Camisia segnis* (Acari: Oribatida). *Intern. J. Acarol.* 10(3), 153-158.

- ARNDT, U., W. NOBEL & U. B. SCHWEIZER, 1987. Bioindikatoren: Möglichkeiten, Grenzen und neue Erkenntnisse. Ulmer, Stuttgart, 388 pp.
- BACHMANN-STEINER, R., R. EGGLI & CH. STRÄHL, 1983. Luftbelastung in der Stadt Zürich. Gesundheitsinspektorat der Stadt Zürich, 18 pp.
- BARTOS, E., 1950. Additions to knowledge of moss-dwelling fauna of Switzerland. Hydrobiologia 2, 285-295.
- BENGTSON, S. A., A. FJELLBERG & T. SOLHÖY, 1974. Abundance of tundra arthropods in Spitsbergen. Ent. Scand. 5, 137-142.
- BERTRAND, M., 1975. Les biotopes des tardigrades "terrestres" dans une hêtraie du massif de l'Aigoual (Cévennes Méridionales). Vie et Milieu 25, 299-314.
- BIERI, M., V. DELUCCHI & C. LIENHARD, 1978. Ein abgeänderter Macfadyen-Apparat für die dynamische Extraktion von Bodenarthropoden. *Mitt. Schweiz. Ent. Ges.* 51, 327-330.
- BLEULER, P. & J. B. BUCHER, 1984. Luftbelastung im Raume Birmensdorf (ZH). Schweiz. Z. Forstwes. 135(9), 801-805.
- BRAUNS, A., 1976. Taschenbuch der Waldinsekten. Bd. 1 u. 2. Georg Fischer, Stuttgart, 817 pp.
- CAIRNS, J. JR., 1983. Are single species toxicity tests alone adequate for estimating environmental hazard? *Hydrobiologia* 100, 47-57.
- CHRISTEN, A. A., 1975. Some fungi associated with Collembola. Rev. Ecol. du Sol 12, 723-728.
- COCHRAN, W. G., 1963. Sampling techniques, 2nd ed. Wiley, New York.
- DAVIS, R. C., 1981. Structure and function of two Antarctic terrestrial moss communities. *Ecol. Monographs* 51(2), 125-143.
- DIAMOND, J., 1986. Overview: Laboratory experiments, field experiments, and natural experiments. *In:* Community Ecology, Diamond J. and T. J. Case (eds.). *Harper and Row*, *New York*, 3-22.
- DONNER, J., 1965. Ordnung Bdelloidea. Akademieverlag, Berlin, 297 pp.
- DUNGER, W., 1983. Tiere im Boden. A. Ziemsen Verlag. Wittenberg Lutherstadt, 280 pp.
- FRAHM, J. P. & W. FREY, 1983. Moosflora. Eugen Ulmer, Stuttgart, 522 pp.
- GERSON, U., 1969. Moss-arthropod associations. Bryologist 72(4), 495-500.
- GERSON, U., 1972. Mites of the genus *Ledermuelleria* (Prostigmata: Stigmaeidae) associated with mosses in Canada. *Acarologia* 13, 319-343.
- GERSON, U., 1982. Bryophytes and invertebrates. In: A. J. E. Smith (ed.), Bryophyte Ecology. Chapmann & Hall, London and New York, 291-332.
- GERSON, U. & M. R. D. SEAWARD, 1977. Lichen-invertebrate associations. *In:* Seaward, M. R. D. (ed.), Lichen Ecology. *Academic Press, London*, 69-119.
- GILBERT, O. L., 1968. Bryophytes as indicators of air pollution in the Tyne Valley. *New Phytol.* 67, 15-30.
- GILBERT, O. L., 1971. Some indirect effects of air pollution on barkliving invertebrates. J. Appl. Ecol. 8, 77-84.
- GISIN, H., 1943. Ökologie und Lebensgemeinschaften der Collembolen im schweizerischen Exkursionsgebiet Basels. Rev. Suisse Zool. 50, 131-224.
- HALE, W. G., 1967. Collembola. In: Burges, A. & F. Raw (eds.), Soil biology. Academic Press, London and New York, 397-411.
- HECK, W. W. & C. S. BRANDT, 1977. Effects of air pollution on vegetation. In: Air Pollution. Vol. II, Stern, A. C. (ed.), Academic Press, New York, 157-229.
- HEINIS, F., 1908. Tardigraden der Schweiz. Zool. Anz. 32, 633-638.
- HEINIS, F., 1910. Systematik und Biologie der moosbewohnenden Rhizopoden, Rotatorien und Tardigraden der Umgebung von Basel. Dissertation Universität Basel.
- HUTER, C., 1986. Schadstoffimmissionen in Wohn- und Erholungsgebieten. Dissertation ETH Zürich, No. 8097, 121 pp.

- HYMAN, L. H., 1951. The Invertebrates: Acanthocephala, Aschelminthes and Entoprocta; the Pseudocoelomate Bilateria. Vol. III. *Mc Graw-Hill, New York*, 572 pp.
- JALIL, M., 1972. The effect of desiccation on some oribatid mites. Proc. Ent. Soc. Wash. 74(4), 406-410.
- KATHMAN, R. D. & S. F. CROSS, 1991. Ecological distribution of moss-dwelling tardigrades on Vancouver Island, British Columbia, Canada. *Can. J. Zool.* 69, 122-129.
- KNÜLLE, W., 1954. Die Arten der Gattung *Tectocepheus* Berlese (Acarina: Oribatei). Zool. Anz. 152, 280-305.
- KOLKWITZ, R., 1959. Ökologie der Saprobien. In: Schriftenr. Ver. Wasser-, Boden- und Lufthygiene 4, 1-64.
- KRANTZ, G. W. (Ed.), 1978. A manual of acarology (Second edition). Oregon State University Book Stores Inc., Corvallis, 509 pp.
- LE BLANC, F. & J. DE SLOOVER, 1970. Relation between industrialization and the distribution and growth of epiphytic lichens and mosses in Montreal. *Can. J. Bot.* 48, 1485-1496.
- LEBRUN, P., 1976. Effets écologiques de la pollution atmosphérique sur les populations et communautés de microarthropodes corticoles (Acariens-Collemboles-Ptérygotes). *Bull. Ecol.* 7, 417-430.
- MAUCCI, W., 1986: Tardigrada. Fauna d'Italia; vol. 24. Calderini, Bologna, 388 pp.
- MCINNES, S. J., 1991. Notes on tardigrades from the Pyrenees, including one new species. *Pedobiologia* 35, 11-26.
- MORGAN, C., 1977. Population dynamics of two species of Tardigrada, *Macrobiotus hufelandi* (Schultze) and *Echiniscus (Echiniscus) testudo* (Doyère), in roof moss from Swansea. J. *Anim. Ecol.* 46, 263-279.
- MUKAMMAL, E., 1976. Review of present knowledge of plant injury by air pollution. *In:* World Meteorol. Org., Technical Note 147, 27 pp.
- MURPHY, P. W. & M. JALIL, 1964. Some observations on the genus *Tectocepheus*. Acarologia, fasc. h.s. (1er Congrès Int. d'Acarologie, Fort Collins, Col., USA, 1963), 187-197.
- NICHOLAS, W. L., 1975. The biology of free-living nematodes, *Clarendon Press*, *Oxford*, 219 pp.
- NIELSEN, C. O., 1967. Nematoda. In: Soil biology. Burges, A. & F. Raw (eds), Academic Press, London, New York, 197-211.
- OKKERMAN, P. C., E. J. V. D. PLASSCHE, W. SLOOFF, C. J. VAN LEEUWEN, & J. H. CANTON, 1991. Ecotoxicological effects assessment: A comparison of several extrapolation procedures. *Ecotoxicol. Environ. Saf.* 8, 254-74.
- PÉREZ-IÑIGO, E., 1975. Contribución al conocimiento de los oribátidos musicolas de la Sierra de Guadarrama y de los Montes de Toledo. Parte II (Acari, Oribatei) *Eos, Madrid* 48, 213-246.
- PETERSEN, H. & M. LUXTON, 1982. A comparative analysis of soil fauna populations and their role in decomposition process. *Oikos* 39, 287-388.
- PSCHORN-WALCHER, H. & P. GUNHOLD, 1957. Zur Kenntnis der Tiergemeinschaft im Moosund Flechtenrasen an Park- und Waldbäumen. Z. Morphol. u. Ökol. Tiere 46, 342-354.
- RAMAZZOTTI, G., 1972. Il Phylum Tardigrada. Mem. Ist. Ital. Idrobiol. Dott. Marco de Marchi 28, 732 pp.
- SANTOS, P. F., J. PHILLIPS & W. G. WHITFORD, 1981. The role of mites and nematodes in early stages of buried litter decomposition in a desert. *Ecology* 62(3), 664-669.
- SCHIEMER, F., 1982. Food dependence and energetics of freeliving nematodes. 1. Respiration, growth and reproduction of *Caenorhabditis briggsae* (Nematoda) at different levels of food supply. *Oecologia* (Berl.) 54, 108-121.
- SCHUBERT, R., 1985. Bioindikation in terrestrischen Ökosystemen. Verlag Fischer Stuttgart, 327 pp.

- SCHWEIZER, J., 1949. Die Landmilben des schweizerischen Nationalparkes. 1. Teil: Parasitiformes Reuter 1909. Lüdin AG, Liestal, 99 pp.
- SCHWEIZER, J., 1951. Landmilben 2. Teil: Trombidiformes. Ergebnisse der wissenschaftlichen Untersuchungen im schweizerischen Nationalpark 3(23), 51-172.
- SCHWEIZER, J., 1956. Landmilben 3. Teil: Sarcoptiformes. Ergebnisse der wissenschaftlichen Untersuchungen im schweizerischen Nationalpark 5(34), 215-377.
- SCHWEIZER, J. & C. BADER, 1963. Die Landmilben der Schweiz (Mittelland, Jura und Alpen). Trombidiformes. Denkschr. Schweiz. Naturf. Gesellsch. 84, 209-378.
- SEAWARD, M. R. D., 1979. Lower plants and the urban landscape. Urban Ecology 4, 217-225.
- SEINHORST, J. W., 1959. A rapid method for the transfer of nematodes from fixative to anhydrous glycerin. *Nematologica* 4, 67-69.
- SMRZ, J., 1992. The ecology of the microarthropod community inhabiting the moss cover of roofs. *Pedobiologia* 36, 331-340.
- STAFANSKI, W., 1923. Etudes sur les nématodes muscicoles des environs de Zakopane. Bull. Acad. Polonaise Sc. Sér. B 1(10), 21-60.
- STEINER, W. A., 1989. Methoden zur Klassifikation der Juvenilstadien einiger Oribatulidae-Arten. Acarologia 30(1), 67-79.
- STEINER, W. A., 1990. The influence of air pollution on moss-dwelling animals. *Dissertation ETH Zürich*, No. 9144, 200 pp.
- STEINER, W. A., 1994. The influence of air pollution on moss-dwelling animals. 2. Aquatic fauna with emphasis on Nematoda and Tardigrada. *Rev. Suisse Zool.* 101(3), in press.
- STEINER, W. A., 1995a. The influence of air pollution on moss-dwelling animals. 3. Terrestrial fauna with emphasis on Oribatida and Collembola. *Acarologia* 36(2), in press.
- STEINER, W. A., 1995b. The influence of air pollution on moss-dwelling animals. 4. Seasonal and long-term fluctuations of rotifer, nematode and tardigrade populations. *Rev. Suisse Zool.* 102(1), in press.
- STEINER, W. A., 1995c. The influence of air pollution on moss-dwelling animals. 5. Fumigation with SO<sub>2</sub> and exposure expreiments. (in prep.).
- STRENZKE, K., 1952. Untersuchungen über die Tiergemeinschaften des Bodens. Die Oribatiden und ihre Synusien in den Böden Norddeutschlands. Zoologica, Stuttgart 37, 1-172.
- TRAVÉ, J., 1963. Ecologie et biologie des Oribates (Acariens) saxicoles et arboricoles. Suppl. 14 à "Vie et Milieu", 267 pp.
- VISSER, S., 1985. Role of the soil invertebrates in determining the composition of soil microbial communities. *In:* Fitter, A. H. (ed.), Ecological interactions in soil. *Blackwell Scientific Publications*, 297-317.
- VON DER DUNK, K., 1988. Das Dach als Lebensraum. 2. Zu den Moosen aufs Dach. Mikrokosmos 77 (10), 300-307.
- WALLWORK, J. A., 1967. Acari. In: Burges, A. & F. Raw (eds.), Soil biology. Academic Press, London and New York, 363-395.
- WEIGMANN, G. & W. KRATZ, 1981. Die deutschen Hornmilbenarten und ihre ökologische Charakteristik. Zool. Beiträge 27(2), 459-489.
- WRIGHT, J. C., 1991. The significance of four xeric parameters in the ecology of terrestrial Tardigrada. J. Zool. Lond. 224, 59-77.

#### APPENDIX: SPECIES LISTS

Total number of individuals: Rotatoria (a); Nematoda (b); Tardigrada (c); Arachnida (Acari excluded), Crustacea, and Myriapoda (d); Oribatida (e); Gamasida and Ixodida (f); Actinedida (g); Collembola (h), "other Insecta" (i) and "all taxa" (k) sampled in tegulous moss cushions of the fumigation and exposure experiments (STEINER 1995c), as well as in epilithic moss cushions of the survey of natural communities (STEINER 1994; STEINER, 1995a), and of the study of seasonal and long-term population dynamics (STEINER 1995b). Since identification to species level was not possible within each taxonomic group, indicated values for " $\Sigma$  Taxa" are minimal values. (n = number of individuals; % = relative number of individuals; - = not analysed; + = less than 0.5%; ult = urban low traffic sites).

(a) Rotatoria		Tegu moss cu		Epilithic moss cushions						
				Survey	y 1984	Survey	v 1987	Dynar	nics	
		n	%	n	%	n	%	n	%	
∑ Rotatoria	$\Sigma$ Individuals	29796	100	6052	100	-	-	3140	100	
(b) Nematoda										
Monhyster	ida									
Monhysteridae		53	+	15	+	-	-	7	+	
Eumonhystera	•	4	+	61	+	-	-			
	f. vulgaris (De Man)	1	+	6	+	-	_ 1	113	1	
Geomonhystera				293	2	-	-			
	australis (Cobb)	157	+	792	4	-	_			
	villosa (Bütschli)	695	2	126	1	-	_	1748	16	
Theristus	(Dationit)	0,2	-	120	•	-	_ ·	1	+	
	entiniae Andrássy			3	+	-	-	-		
Araeolaimi	i d a	2		5						
Plectidae		3	+	5	+	-		1.4		
	ulosus (Bastian)			32	+	-	-	14	+	
	matus (Bütschli)	2174	0	6	+	-	-	1	+	
	inatus Bastian s.l.	3174	8	82	+	-	-	1100	10	
Plectus cf. parie		569	1	384	2		-	1102	10	
Plectus parvus B	astian	663	2	315	2	-	-	203	2	
	undetermined)"	9122	24	1994	10	-	-	2995	27	
	andrassyi (Timm)	13276	34	590	3	-	-	2409	22	
Tylocephalus au	riculatus (Bütschli)	1836	5	882	5	-	-	255	2	
Chromador										
Prodesmodora te	erricola Altherr			4	+	-	-			
Rhabditida										
Rhabditida sp. 1				1	+		-			
	terrestris (Bütschli)			2	+	-	-	1	+	
Eucephalobus		1	+			-	-			
Eucephalobus st	riatus (Bastian)	3	+			~	-			
Cepĥalobus				8	+	-	-			
Acrobeloides				2	+	-	-			
Ypsilonellus						-	-	12	+	
	imucronatus (Sumenko	va) 5	+	4	+	-	-			
	illiger (De Man)	-				-	-	1	+	
Panagrolaimus	0	18	+	291	2	-	-	115	1	
	cf. rigidus (Schneider)	161	+	10	+	-	-	7	+	
	cf. rigidus (Schneider)			107	1	-	_	19	+	
Protorhabditina				1	+	_	-			
Protorhabditis s				27	+	_	_			
Mesorhabditina				2	+		-			
	hystera (Bütschli)			$\frac{2}{2}$	+	-	-			
Tylenchida		1	+	7	+			1	+	
Tylenchida sp. 1		1	Ŧ			-	-	1	+	
i viencniaa sp. 1				1	+	-	-			

(b) Nematoda (continued)	Tegul moss cus		Epilithic moss cushions						
	n	%	Survey n	1984 %	Surve	y 1987 %	Dyr n	amics %	
		/0	11	/0		/0		70	
Tylenchidae	7	+			-	-			
Tylenchus arcuatus Siddiqi			1	+		-			
Tylenchus elegans De Man			586	3	-	-			
Filenchus			5	+	-	-			
Haplolaimidae	2	+			-	-	10	+	
Neotylenchidae			1	+	-	-			
Neotylenchidae sp. 1 larvae			209	1	-	-			
Neotylenchiade sp. 2 larvae			24	+	~	-			
Neotylenchidae sp. 3			16	+	-	-			
Neotylenchidae sp. 4			19	+	-	-			
Aphelenchoides	19	+	68	+	-	-	14	+	
Aphelenchoides sp. 1	240	1	72	+	-	-	8	+	
Aphelenchoides sp. 2			32	+	-	-			
Aphelenchoides sp. 3	17	+			-	-	3	+	
Aphelenchoides sp. 4	31	+	2694	14	-	-	15	+	
Aphelenchoides sp. 5	462	1	6736	35	-	-	633	6	
Aphelenchoides sp. 6			507	3	-	-			
Laimaphelenchus	3	+	374	2	-	*	1	+	
Laimaphelenchus sp. 1									
(gr. pannocaudus Massey)			254	1	-	-			
Laimaphelenchus deconincki									
Elmiligy & Geraert			149	1	-	-	1	+	
Enoplida									
Paratripyla intermedia (Bütschli)	6548	17	443	2	-	-	922	8	
Dorylaimida									
Mononchidae	49	+	429	2	-	-			
Prionchulus muscorum (Dujardin)	956	2	144	1	-	-	63	1	
Nygolaimus sp. 1	0		81	1	-	-	16		
Dorylaimidae	2	+	3	+	-	-	16	+	
Dorylaimidae sp. 1			1	+	-	-			
Dorylaimidae sp. 2			45	+	-	-			
Eudorylaimus sp. 1			1	+	-	-			
Eudorylaimus sp. 2			53	+	-	-			
Eudorylaimus sp. 3			6	+	-	-			
Eudorylaimus sp. (gr. carteri [Bastian])					-	-	58	1	
Eudorylaimus sp. (gr. iners [Bastian])					-	-	71	1	
Mesodorylaimus sp. 1	19	+	1	+	-	-	_		
Mesodorylaimus sp. 2					-	-	6	+	
Mesodorylaimus sp. 3					-	-	9	+	
Labronema sp. 1					~	-	79	1	
Nematoda (undetermined)	464	1	45	+	-	-	22	+	
$\sum \text{Nematoda} \qquad \sum \text{Individuals} \\ \sum \text{Taxa}$	38561 21	100	19054 47	100	-	-	10935 26	100	
Nematoda treated as a group (ult)			12350						
(c) Tardigrada									
Heterotardigrada									
Echiniscus blumi Richters Echiniscus testudo (Doyère)	6790	8	11	+	-	-			
	146	+	887	10					

(c) Tardigrada (continued)	Tegul moss cu		Εŗ	oilith	nic ma	oss cu	ushions		
	n	%	Survey n	1984 %	Survey n	1987 %	Dyna n	mics %	
Echiniscus trisetosus Cuénot Echiniscus	16	. +	42 1	+ +	-				
Eutardigrada	100.00		100						
Macrobiotus areolatus Murray Macrobiotus cf. artipharyngis Iharos	19829 16254	24 19	192	2	-	-			
Macrobiolus cj. artipharyngis maros Macrobiotus hufelandi Schultze	27535	33	2880	33	-	-	202	9	
Macrobiotus persimilis Binda & Pilate		1	2177	25	-	-	722	33	
Macrobiotus richtersi Murray			1	+	-	-	33	1	
Macrobiotus	1972	2	533	6	-	-	16	1	
Hypsibius cf. bakonyiensis (Iharos)			29	+	-	-			
Hypsibius convergens Urbanowicz s.l.		4	132	2	-	-	291	13	
Hypsibius oberhäuseri (Doyère)	6490	8	933	11	-	-	921	42	
Hypsibius cf. pallidus Thulin	22	+	8	+	-	-	12	1	
Hypsibius	139	+	16	+	-	-	8	+	
Diphascon pingue (Marcus) -	195	+			-	-			
Ishypsibius prosostomus (Thulin)	30	+	279 512	3	-	-	10	+	
Milnesium tardigradum Doyère	206	+	312	0					
Eutardigrada (undetermined)	9	+	3	+	-	-			
$\Sigma$ Tardigrada $\sum_{\Sigma}$ Individuals $\sum_{\Sigma}$ Taxa	83612 12	100	8636 13	100	-	-	2215 7	100	
Tardigrada treated as a group (ult)			2706						
(d) Arachnida (Acari excluded) Crustacea, and Myriapoda									
Pseudoscorpiones									
Chthonius			1	12					
Araneae									
Theridiidae			1	12					
Micryphantidae			2 2	25					
Linyphildae			$\frac{2}{1}$	25 12					
Agelenidae			1	12					
O p i l i o n e s Phalangiidae			1	12					
$\Sigma$ Arachnida $\Sigma$ Individuals			8	100					
$\sum \text{ At a cliffida}$ $\sum \text{ Taxa}$			6	100					
I s o p o d a Oniscidae			24	100					
$\sum Crustacea \qquad \sum Individuals \\ \sum Taxa$			24 1	100					
C h i l o p o d a Lithobiidae Geophilidae			4 7	4 7			1	17	
D i p l o p o d a Polyxenus lagurus L. Blaniulidae	1		93 1	88 1	29	100	8	83	

#### WERNER A. STEINER

(d) Arachnida (continued)	r	Tegu noss ci	llous ushions	Epilithic moss cushio					
		n	%	Survey n	1984 %	Survey n	1987 %	Dyr n	amics %
Pauropoda			-	. 1	1				
Σ Myriapoda Σ Indivio Σ Taxa	duals	1 1	100	106 5	100	29 1	100	6 2	100
(e) Oribatida Brachychthoniidae Liochthonius Verachthonius laticeps (Stren	uzke)	2	+	2 10	+ +	2	+		
Camisia segnis (Hermann) Trhypochthonius tectorum Be Damaeus	erlese	283	30	360 221 3	4 2 +	111 3	6 +	1 16	+ 5
Eremaeus oblongus Koch Tectocepheus sarekensis Träg	gårdh	197	21	737 5866	7 57	4 1126	+ 60	112	37
Oppiidae Oppia nitens (Koch) Opiella nova (Oudemans) Quadroppia quadricarinata ( Oppia fasciata (Paoli) Suctobelba sarekensis Forssh				8 3 8 1 2	+ + + +	1	+	114	38
<i>Cymberemaeus cymba</i> (Nicol <i>Micreremus brevipes</i> (Michae <i>Scutovertex cf. sculptus</i> Mich	ël)	1 1 292	+ + 31	8 36	+ +	5 1 4	+ + +		
Dometorina plantivaga (Berl Oribatula tibialis (Nicolet) Phauloppia lucorum (Koch) Phauloppia paspalevi Csiszas Scheloribates laevigatus (Koc	r	18	2	1 132 57 1 23	+ 1 1 + +	131 170	7 9	2	1
Scheloribates latipes (Koch) Zygoribatula exilis (Nicolet) Zygoribatula frisiae (Oudema Zygoribatula propinquus (Ou	ans)	9 6 5	1 1 1	410 1903 15 58	4 19 + 1	17 78 20	1 4 1	27	9
Chamobates sp. 1 Chamobates borealis Trägård	lh			3 36	+++	20		10	3
Ceratozetes cf. minutissimus Trichoribates cf. trimaculatus Mycobates parmeliae (Micha Minunthozetes pseudofusiger Minunthozetes semirufus (Koc Punctoribates punctum (Koch	s (Koch) ël) Schweizer ch)	92 2	10 +	1 213 136 5 6	+ 2 1 + +	193	10	10	3
Protoribates cf. capucinus Be Eupelops planicornis (Koch) Oribatella quadricornuta (M Galumna alatus (Hermann)				3 8	+ +	1 1	+ +		
Oribatida sp. 1 Oribatida sp. 2				1 1	+ +				
Oribatida (undetermined)		45	5					9	3
$ \sum \text{Oribatica} \qquad \sum \text{Indivis} \\ \sum \text{Taxa} $	duals	953 12	100	10278 33	100	1868 17	100	301 8	100

.

(f) Gamasida and Ixodida	Teg moss	gulous cushions	Е	pilith	ic mo	ss cus	hion	S
	n	%	Surve	ey 1984 %	Survey n	1987 %	Dyn n	amics %
G a m a s i d a Laelaptidae			5	5				
Hypoaspis Hypoaspis claviger Karg	1	14	3	3				
Phytoseiidae	2	29	. <u>.</u>		1	11		
Amblyseiulus Amblyseiulus murteri (Schweizer)	1	14	5	5	1	11		
Typhlodromus Typhlodromus andrei Karg	1	14	9	8				
Lasioseius fimetorum Karg		-	3	3		·		
Halolaelaps Asca aphioides (L.)			$\frac{1}{33}$	1 30	4	44		
Arctoseius cetratus Sellnik			2	2				
Zerconidae Zercon montanus Willmann			3 3	3 3				
Rhodacaridae Dendrolaelaps			2 6	2 5	2	22		
Gamasellus falciger (Can.)	1	14			-			
Parasitidae Pergamasus crassipes (L.)			4 6	4 5				
Gamasina (undetermined)	1	14	12	11	1	11		
Uropodidae Uroobovella notabilis Berlese			6 4	5 4				
I x o d i d a Ixodidae			3	3				
$\sum Gamasida + Ixodida \sum_{\sum Taxa} Individuals$	7 4	100	110 12	100	9 3	100	0 0	
(g) Actinedida Pachygnathidae			1	4				
Nanorchestidae	1	4	22	+2	6	2		
Eupodes Penthalodidae			454 1	36 +	71	27	4	8
Penthaleidae			70	6				
Paratydaeolus	2	11	12	1	50		11	
Tydeidae Triophtydeus	3	11	$\begin{array}{c} 71 \\ 10 \end{array}$	6 1	59	23	11	23
Microtydeus			3	+				
Tydeus bedfordiensis Evans			14	1				
Tydeus (type Lorryia 2) Tydeus (Type Paralorryia 1)			12 9	1 1				
Tydeus (Type Paralorryia 2) Tydeus (Type Tydeus sp. V)			7 32	1 3				
Tydeus (Type Tydeus sp. A)			23	2				
Bdellidae Bdella longicornis (L.)	2	7	1 103	+ 8	17	7	17	35
Cyta			65	5	2	1	1	2
Spinibdellinae Cunaxidae	1 4	4 15	2 13	+ 1 1	26	10		5
Cunaxoides			8	1				

#### WERNER A. STEINER

# (g) Actinedida

## Tegulous moss cushions E pilithic moss cushions

		moss c	usmons	L	piiiti			SHIOHS		
		n	%	Survey n	y 1984 %	Surve	y 1987 %	Dy: n	namics %	
Pygmephoroidea				2						
Pygmephoridae	1			. 3 . 4	++					
Scutcaridae				1	+					
Tarsonemidae		1	4	289	23	1	+	1	2	
Raphignathidae		1	4	2	+					
Cryptognathidae	;					2	1	4	8	
Caligonellidae				5		. 2	1			
Stigmaeidae		. <u></u>		5	+					
Bryobia		1	4	6	+	6	2			
Tenuipalpidae Eriophidae		1 1	4 4							
Erythraeidae		5	19			60	23			
Johnstonionidae		5	17			2	1			
Podotrombidiida				1	+					
Trombidiidae		1	4	4	+	1	+			
Actinedida (und	etermined)	6	22	8	1	4	2	10	21	
$\sum$ Actinedida	$\Sigma$ Individuals	27	100	1256	100	259	100	48	100	
-	$\overline{\Sigma}$ Taxa	11		29		13		6		
(h) Collembola										
Arthropleo	na									
Hypogastrura				16	+					
Hypogastrura sp				5		2	1			
	ssimilis Krausbauer E. elevata Cassagnau			5 3	+ +					
	igadinensis Gisin			1	+					
Hypogastrura tu				1		25	12	47	73	
Hypogastrura ve				74	2	2	1			
Schoettella unur	guiculata (Tullberg)			4047	88	52	25			
Xenylla	- -			6	+					
Xenylla boerner				1	+					
Xenylla brevisin		1130	95	43	1	55	26	11	17	
Xenylla welchi H Willemia anophi				11	+					
Friesea claviset				31	+ 1	1	+			
	parvula (Schäffer)			4	+		·			
Neanura muscoi						1	+			
Onychiuridae				4	+	2	1			
Mesaphorura				6	+					
Mesaphorura kr	ausbaueri Börner			20	+					
Folsomides ang		18	2							
Folsomia similis	Bagnall			3	+					
Isotomiella para				2	+	1				
Proisotoma han				10		1	+			
Isotoma arborea Isotoma viridis 1				18 1	++					
	ibilis (Schäffer)			29	1	5	2			
Parisotoma not				-		5	2			
					+					
Parisotoma nota Isotomurus plun Isotomurus palu	iosus Bagnall			1 6	++	1	+			

(h) Collembola (continued)	Tegu moss cu		E	pilith	ic me	oss cu	shio	n s
	n	%	Surve n	ey 1984 %	Survey 1987 n %		Dy: n	namics %
Entomobrya Entomobrya multifasciata (Tullberg) Entomobrya nivalis (L.) Entomobrya marginata (Tullberg) Orchesella Orchesella cincta (L.) Willowsia buski (Lubbock) Willowsia nigromaculata (Lubbock)			$   \begin{array}{r}     16 \\     97 \\     16 \\     61 \\     1 \\     25 \\     35 \\     3 \\     3   \end{array} $	+ 2 + 1 + 1 1 + 1 +	2 20 20 8 4		3	5
Lepidocyrtus Lepidocyrtus gr. servicus-pallidus Lepidocyrtus lignorum (Fabricius) Lepidocystus cyaneus Tullberg Pseudosinella Pseudosinella sp. 1 Pseudosinella-Lepidocyrtus			2 1 12 4 2	+ + + + +	1	+		
S y m p h y p l e o n a Sminthurinus Sminthurinus aureus (Lubbock) Sminthurinus bimaculatus (Axelson) Bourletiella	10	1	1 3	+ +	1 4 1	+ 2 +		
Collembola (undetermined)	36	3	2	+			3	5
$\sum Collembola \qquad \sum Individuals \\ \sum Taxa$	1194 3	100	4617 32	100	208 18	100	64 3	100
(i) Insecta (Collembola excluded) D i p l u r a			2	I				
Psocoptera	4	10	7	4				
Thysanoptera			6	3				
Heteroptera Anthocoridae			2 1	1 1	1	3		
H o m o p t e r a Coccina Aleyrodina Aphidina			4 5 4	2 3 2				
C o l e o p t e r a Staphylinidae Curculionidae Ptiliidae	1	3	2 4	1 2	2 1 1	5 3 3		
N e u r o p t e r a Chrysopa Coniopterygidae			1	1	1	3		
H y m e n o p t e r a Mymaridae Braconidae Scelionidae	3	8	1 8 1 2	1 4 1 1				
D i p t e r a Chironomidae	11 19	28 49	16 41	8 21	2 20	5 50	3	75

(i) Insecta (continued)		ilous ushions	Epilithic moss cushions						
			Surve	Survey 1984		y 1987	Dynamic		
	n	%	n	%	n	%	n	%	
Psychodidae			. 32	. 17	4	10			
Cedicomyiidae			17	9	3	8 3			
Sciaridae	1	3	20	10	1	3			
Scatopsidae			1	1					
Drosophilidae			1	1					
Trichoptera									
Hydroptila sparsa Curtis			2	1					
Hydroptila pulchicornis Pictet			1	1					
Lepidoptera									
Lepidoptera (larvae)			9	5	2	5	1	25	
Insecta (undetermined)			1	1	2	5			
$\Sigma$ Insecta $\Sigma$ Individuals	39	100	191	100	40	100	4	100	
∑ Taxa	5		21		10		2		

(k) All taxa		Tegulous moss cushions	Epilithic moss cushions			Total
		moss cusmons	Survey 1984	Survey 1987	Dynamics	
$\sum$ Aquatic fauna	$\sum_{\Sigma \text{ Individuals}} \text{Individuals}$	151'969 34	48'798 61		16'290 34	217'057 72
$\Sigma$ Terrestrial fauna	$\frac{\sum \text{Individuals}}{\sum \text{Taxa}}$	2'221 36	13'103 133	2'413 62	423 21	21'647 162
$\Sigma$ Aquatic + terrestrial fauna	$\frac{\sum \text{Individuals}}{\sum \text{Taxa}}$	154'190 70	6'421 194	2'413 62	16'713 55	238'704 234